

# جداول میکروارگانیزم های بیماریزای اولویت دار و آنتی بیوتیک های تعیین شده برای آزمایش تعیین حساسیت ضد میکروبی در برنامه مهار مقاومت میکروبی

ویرایش ششم

بر اساس CLSI M100 31<sup>th</sup> ed., 2021

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آزمایشگاه مرجع سلامت

وزارت بهداشت، درمان و آموزش پزشکی

۱۴۰۰

<b><i>Escherichia coli</i></b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>PENICILLINS</b>					
Ampicillin	10 µg	≥ 17	14–16 <sup>^</sup>	≤ 13	Results of ampicillin testing can be used to predict results for amoxicillin.
<b>CEPHEMS</b>					
Cefazolin (PARENTERAL)	30 µg	≥ 23	20–22	≤ 19	Breakpoints when cefazolin is used for therapy of infections other than uncomplicated UTIs due to <i>E.coli</i> , <i>K. pneumoniae</i> & <i>P.mirabilis</i> . Breakpoints are based on a dosage regimen of 2 g administered every 8 h.
Cefazolin (PARENTERAL) (urine)	30 µg	≥ 15	-	≤ 14	Breakpoints when cefazolin is used for therapy of uncomplicated UTIs due to <i>E.coli</i> , <i>K pneumoniae</i> & <i>P.mirabilis</i> . Breakpoints are based on a dosage regimen of 1 g administered every 12 h.
Cefazolin (ORAL) (surrogate test for oral cephalosporins & uncomplicated UTI) (urine)	30 µg	≥ 15	-	≤ 14	(a) Breakpoints are for cefazolin when used as a surrogate test to predict results for the oral agents cefaclor, cefdinir, cefpodoxime, cefprozil, cefuroxime, cephalixin, and loracarbef when used for therapy of uncomplicated UTIs due to <i>E. coli</i> , <i>K. pneumoniae</i> , and <i>P. mirabilis</i> . Cefazolin as a surrogate may overcall resistance to cefdinir, cefpodoxime, and cefuroxime. If cefazolin tests resistant, test these drugs individually if needed for therapy.
Cefepime	30 µg	≥ 25	19–24	≤ 18	The breakpoint for susceptible is based on a dosage regimen of 1 g administered every 12 h. The Breakpoint for SDD is based on dosing regimens that result in higher cefepime exposure, either higher doses or more frequent doses or both, up to approved maximum dosing regimens. *SDD: Susceptible-Dose Dependent



الوزارة  
الصحة  
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<b>Escherichia coli (continued)</b>											
Cefotaxime or Ceftriaxone	30 µg 30 µg	≥ 26 ≥ 23	23–25^ 20–22^	≤ 22 ≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 24 h for ceftriaxone and 1 g administered every 8 h for cefotaxime.						
Ceftazidime	30 µg	≥ 21	18–20^	≤ 17	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.						
<b>CARBAPENEMS</b>											
Imipenem	10 µg	≥ 23	20–22^	≤ 19	Breakpoints are based on a dosage regimen of 500 mg administered every 6 h or 1 g every 8 h.						
Meropenem	10 µg	≥ 23	20–22^	≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.						
<b>LIPOPEPTIDES</b>											
Colistin or Polymixin B		-	-	-	(a) Colistin (methanesulfonate) should be given with a loading dose and maximum renally adjusted dose. (b) Polymixin B should be given with a loading dose and maximum recommended dose. (c) When colistin or polymixin B is given systemically, neither is likely to be effective for pneumonia. (d) For colistin, broth microdilution, CBDE, and CAT MIC methods are acceptable. For polymixin B, broth microdilution is the only approved method. Disk diffusion and gradient diffusion methods should not be performed (see Table 3D, Page 142-147). *CAT: Colistin Agar Test *CBDE: Colistin Broth Disk Elution						
					<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>						
					<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 33%;">S</th> <th style="width: 33%;">I</th> <th style="width: 33%;">R</th> </tr> </thead> <tbody> <tr> <td style="text-align: center;">-</td> <td style="text-align: center;">≤ 2</td> <td style="text-align: center;">≥ 4</td> </tr> </tbody> </table>	S	I	R	-	≤ 2	≥ 4
S	I	R									
-	≤ 2	≥ 4									
<b>AMINOGLYCOSIDES</b>											
Gentamicin	10 µg	≥ 15	13-14^	≤ 12							
Amikacin	30 µg	≥ 17	15–16^	≤ 14							
<b>FLUOROQUINOLONES</b>											
Ciprofloxacin	5 µg	≥ 26	22-25^	≤ 21	Breakpoints for ciprofloxacin are based on a dosage regimen of 400 mg IV or 500 mg orally administered every 12 h.						

<b><i>Escherichia coli</i> (continued)</b>					
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim- sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11–15	≤ 10	
<b>NITROFURANS</b>					
Nitrofurantoin	300 µg	≥ 17	15–16	≤ 14	For testing and reporting urinary tract isolates only.



وزارت صحت  
حکومت سندھ

<b><i>Klebsiella pneumoniae</i></b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>CEPHEMS</b>					
Cefazolin (PARENTERAL)	30 µg	≥ 23	20–22	≤ 19	Breakpoints when cefazolin is used for therapy of infections other than uncomplicated UTIs due to <i>E.coli</i> , <i>K pneumoniae</i> & <i>P.mirabilis</i> . Breakpoints are based on a dosage regimen of 2 g administered every 8 h.
Cefazolin (PARENTERAL) (urine)	30 µg	≥ 15	-	≤ 14	Breakpoints when cefazolin is used for therapy of uncomplicated UTIs due to <i>E.coli</i> , <i>K pneumoniae</i> & <i>P.mirabilis</i> . Breakpoints are based on a dosage regimen of 1 g administered every 12 h.
Cefazolin (ORAL) (surrogate test for oral cephalosporins & uncomplicated UTI) (urine)	30 µg	≥ 15	-	≤ 14	(a) Breakpoints are for cefazolin when cefazolin results are used to predict results for the oral agents cefaclor, cefdinir, cefpodoxime, cefprozil, cefuroxime, cephalexin, and loracarbef when used for therapy of uncomplicated UTIs due to <i>E. coli</i> , <i>K. pneumoniae</i> , and <i>P. mirabilis</i> . Cefazolin as a surrogate may overcall resistance to cefdinir, cefpodoxime, and cefuroxime. If cefazolin tests resistant, test these drugs individually if needed for therapy.
Cefepime	30 µg	≥ 25	19–24	≤ 18	The Breakpoint for susceptible is based on a dosage regimen of 1 g every 12 h. The Breakpoint for SDD* is based on dosing regimens that result in higher cefepime exposure, either higher doses or more frequent doses or both, up to approved maximum dosing regimens. *SDD: Susceptible-Dose Dependent
Cefotaxime or Ceftriaxone	30 µg 30 µg	≥ 26 ≥ 23	23–25^ 20–22^	≤ 22 ≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 24 h for ceftriaxone and 1 g administered every 8 h for cefotaxime.
Ceftazidime	30 µg	≥ 21	18–20^	≤ 17	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.



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وزارة الصحة

<b><i>Klebsiella pneumonia</i> (continued)</b>					
<b>CARBAPENEMS</b>					
Imipenem	10 µg	≥ 23	20–22 <sup>^</sup>	≤ 19	Breakpoints are based on a dosage regimen of 500 mg administered every 6 h or 1 g every 8 h.
Meropenem	10 µg	≥ 23	20–22 <sup>^</sup>	≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.
<b>LIPOPEPTIDES</b>					
Colistin or Polymixin B	-	-	-	-	<p>(a) Colistin (methanesulfonate) should be given with a loading dose and maximum renally adjusted dose.</p> <p>(b) Polymixin B should be given with a loading dose and maximum recommended doses.</p> <p>(c) When colistin or polymixin B is given systemically, neither is likely to be effective for pneumonia.</p> <p>(d) For colistin, broth microdilution, CBDE, and CAT MIC methods are acceptable. For polymixin B, broth microdilution is the only approved method. Disk diffusion and gradient diffusion methods should not be performed (see Table 3D, Page 142-147).</p> <p>*CAT: Colistin Agar Test</p> <p>*CBDE: Colistin Broth Disk Elution</p>
<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>					
<b>S</b>		<b>I</b>		<b>R</b>	
-		≤ 2		≥ 4	
<b>AMINOGLYCOSIDES</b>					
Gentamicin	10 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	
Amikacin	30 µg	≥ 17	15–16 <sup>^</sup>	≤ 14	
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 26	22-25 <sup>^</sup>	≤ 21	Breakpoints for ciprofloxacin are based on a dosage regimen of 400 mg IV or 500 mg orally administered every 12 h.
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11–15	≤ 10	
<b>NITROFURANS</b>					
Nitrofurantoin	300 µg	≥ 17	15–16	≤ 14	For testing and reporting urinary tract isolates only.



\*When fecal isolates of *Salmonella* are tested, only ampicillin, a fluoroquinolone, and trimethoprim-sulfamethoxazole should be reported routinely. In addition, for extraintestinal isolates of *Salmonella* spp., a third-generation cephalosporin and chloramphenicol should be tested and reported.

<b><i>Salmonella</i> spp.</b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>PENICILLINS</b>					
Ampicillin	10 µg	≥ 17	14–16 <sup>^</sup>	≤ 13	Results of ampicillin testing can be used to predict results for amoxicillin.
<b>CEPHEMS</b>					
Ceftriaxone (For extraintestinal isolate)	30 µg	≥ 23	20–22 <sup>^</sup>	≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 24 h for ceftriaxone
Ceftazidime (For extraintestinal isolate)	30 µg	≥ 21	18–20 <sup>^</sup>	≤ 17	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 31	21–30 <sup>^</sup>	≤ 20	Isolates of <i>Salmonella</i> spp. that test not susceptible to ciprofloxacin, levofloxacin, ofloxacin, or pefloxacin may be associated with clinical failure or delayed response in fluoroquinolone-treated patients with salmonellosis.
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11–15	≤ 10	
<b>PHENICOLS</b>					
Chloramphenicol	30 µg	≥ 18	13–17	≤ 12	
<b>MACROLIDS</b>					
Azithromycin	15 µg	≥ 13	-	≤ 12	(a) <i>S. enterica</i> ser. Typhi only: breakpoints are based on MIC distribution data and limited clinical data. <b>(b) Breakpoints are based on a dosage regimen of 500 mg administered daily.</b>



المنشأة الوطنية لمكافحة الأمراض

\*When fecal isolates of *Shigella* spp. are tested, only ampicillin, a fluoroquinolone, and trimethoprim-sulfamethoxazole should be reported routinely.

<b><i>Shigella</i> spp.</b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>PENICILLINS</b>					
Ampicillin	10 µg	≥ 17	14–16 <sup>^</sup>	≤ 13	Results of ampicillin testing can be used to predict results for amoxicillin.
<b>CEPHEMS</b>					
Ceftriaxone (Only for ciprofloxacin resistant strain)	30 µg	≥ 23	20–22 <sup>^</sup>	≤ 19	Breakpoints are based on a dosage regimen of 1 g administered every 24 h for ceftriaxone
Ceftazidime (Only for ciprofloxacin resistant strain)	30 µg	≥ 21	18–20 <sup>^</sup>	≤ 17	Breakpoints are based on a dosage regimen of 1 g administered every 8 h.
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 26	22–25 <sup>^</sup>	≤ 21	Breakpoints for ciprofloxacin are based on a dosage regimen of 400 mg IV or 500 mg orally administered every 12 h.
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11–15	≤ 10	
<b>MACROLIDES</b>					
Azithromycin	15 µg	≥ 16	11–15	≤ 10	(a) <i>Shigella</i> spp. only: azithromycin disk diffusion zones can be hazy and difficult to measure, especially <i>S. sonnei</i> . If an isolate has a zone of inhibition that is difficult to measure, an MIC method is recommended. Media source may affect the clarity of the end points for disk diffusion tests. (b) Breakpoints are based on a dosage regimen of 500 mg administered daily.





## Tests for Extended-Spectrum $\beta$ -Lactamases in *Escherichia coli*, *Klebsiella pneumoniae*, *Salmonella* spp and *Shigella* spp.

Test	Criteria for Performance of ESBL Test	ESBL Test
Antimicrobial concentration	<p>Cefpodoxime 10 <math>\mu</math>g or            Ceftazidime 30 <math>\mu</math>g or            Aztreonam 30 <math>\mu</math>g or            Cefotaxime 30 <math>\mu</math>g or            Ceftriaxone 30 <math>\mu</math>g</p> <p>(Testing more than one antimicrobial agent improves the sensitivity of ESBL detection.)</p>	<p>Ceftazidime 30 <math>\mu</math>g            Ceftazidime-clavulanate 30/10 <math>\mu</math>g</p> <p><u>and</u></p> <p>Cefotaxime 30 <math>\mu</math>g            Cefotaxime-clavulanate 30/10 <math>\mu</math>g</p> <p>(Testing necessitates using both cefotaxime and ceftazidime, alone and in combination with clavulanate.)</p>
Results	<p>Cefpodoxime zone <math>\leq</math> 17 mm            Ceftazidime zone <math>\leq</math> 22 mm            Aztreonam zone <math>\leq</math> 27 mm            Cefotaxime zone <math>\leq</math> 27 mm            Ceftriaxone zone <math>\leq</math> 25 mm</p> <p>Zones above may indicate ESBL production.</p>	<p>A <math>\geq</math> 5mm increase in a zone diameter for either antimicrobial agent tested in combination with clavulanate vs the zone diameter of the agent when tested alone = ESBL (eg, ceftazidime zone = 16; ceftazidime-clavulanate zone = 21).</p>
Reporting		<p>For all confirmed ESBL-producing strains:</p> <p>If laboratories do not use current cephalosporin and aztreonam breakpoints, the test interpretation should be reported as resistant for all penicillins, cephalosporins, and aztreonam.</p> <p>If laboratories use current cephalosporin and aztreonam breakpoints, then test interpretations for these agents do not need to be changed from susceptible to resistant.</p>



الجمهورية العربية السعودية  
وزارة الصحة

<b><i>Pseudomonas aeruginosa</i></b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>β-LACTAM/β-LACTAMASE INHIBITOR COMBINATIONS</b>					
Piperacillin-tazobactam	100/10 µg	≥ 21	15–20 <sup>^</sup>	≤ 14	Breakpoints for piperacillin (alone or with tazobactam) are based on a piperacillin dosage regimen of at least 3 g administered every 6 h.
<b>CEPHEMS</b>					
Cefepime	30 µg	≥ 18	15-17 <sup>^</sup>	≤ 14	Breakpoints are based on a dosage regimen of 1 g administered every 8 h or 2 g administered every 12 h.
Ceftazidime	30 µg	≥ 18	15-17 <sup>^</sup>	≤ 14	Breakpoints are based on a dosage regimen of 1 g administered every 6 h or 2 g administered every 8 h.
<b>LIPOPEPTID</b>					
Colistin or Polymixin B	-	-	-	-	(a) Colistin (methanesulfonate) should be given with a loading dose and maximum renally adjusted doses.
					(b) Polymixin B should be given with a loading dose and maximum recommended doses.
					(c) When colistin or polymixin B is given systemically, neither is likely to be effective for pneumonia.
					(d) For colistin, broth microdilution, CBDE, and CAT MIC methods are acceptable. For polymixin B, broth microdilution is the only approved method. Disk diffusion and gradient diffusion methods should not be performed (see Table 3D, Page 142-147).
					*CAT: Colistin Agar Test
					*CBDE: Colistin Broth Disk Elution
					<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>
		<b>S</b>	<b>I</b>	<b>R</b>	
		-	≤ 2	≥ 4	



الوزارة  
الصحة  
السعودية

<b><i>Pseudomonas aeruginosa</i> (continued)</b>					
<b>CARBAPENEMS</b>					
Imipenem	10 µg	≥ 19	16-18 <sup>^</sup>	≤ 15	Breakpoints for imipenem are based on a dosage regimen of 1 g administered every 8 h or 500 mg administered every 6 h.
Meropenem	10 µg	≥ 19	16-18 <sup>^</sup>	≤ 15	Breakpoints for meropenem are based on a dosage regimen of 1 g administered every 8 h.
<b>AMINOGLYCOSIDES</b>					
Gentamicin	10 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	
Tobramycin	10 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	
Amikacin	30 µg	≥ 17	15-16 <sup>^</sup>	≤ 14	
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 25	19-24 <sup>^</sup>	≤ 18	Breakpoints are based on a dosage regimen of 400 mg IV administered every 8 h.



وزارت صحت سندھ

<b>Acinetobacter spp.</b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>β-LACTAM/β-LACTAMASE INHIBITOR COMBINATIONS</b>					
Ampicillin-sulbactam	10/10 µg	≥ 15	12-14	≤ 11	
Piperacillin-tazobactam	100/10 µg	≥ 21	18-20	≤ 17	
<b>CEPHEMS</b>					
Cefepime	30 µg	≥ 18	15-17	≤ 14	
Ceftazidime	30 µg	≥ 18	15-17	≤ 14	
<b>CARBAPENEMS</b>					
Imipenem	10 µg	≥ 22	19-21	≤ 18	Breakpoints are based on a dosage regimen of 500 mg administered every 6 h.
Meropenem	10 µg	≥ 18	15-17	≤ 14	Breakpoints are based on a dosage regimen of 1 g administered every 8 h or 500 mg administered every 6 h.
<b>LIPOPEPTID</b>					
Colistin or Polymixin B	-	-	-	-	(a) Colistin (methanesulfonate) should be given with a loading dose and maximum renally adjusted doses.
					(b) Polymixin B should be given with a loading dose and maximum recommended doses.
					(c) When colistin or polymixin B is given systemically, the drug is unlikely to be effective for pneumonia.
					(d) The only approved MIC methods is broth microdilution, CBDE, CAT, disk diffusion, and gradient diffusion should not be performed (see Table 3D, Page 142-147). *CAT: Colistin Agar Test *CBDE: Colistin Broth Disk Elution
					<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>
		<b>S</b>	<b>I</b>	<b>R</b>	
		-	≤ 2	≥ 4	



وزارت صحت سندھ

<b><i>Acinetobacter</i> spp. (continued)</b>					
<b>AMINOGLYCOSIDES</b>					
Gentamicin	10 µg	≥ 15	13-14	≤ 12	
Tobramycin	10 µg	≥ 15	13-14	≤ 12	
Amikacin	30 µg	≥ 17	15-16	≤ 14	
<b>TETRACYCLINES</b>					
Minocycline	30 µg	≥ 16	13-15	≤ 12	
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 21	16-20	≤ 15	
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11-15	≤ 10	



انجمن صحت ملی  
حکومت سندھ

<b><i>Staphylococcus aureus</i></b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>PENICILLINASE-LABILE PENICILLINS</b>					
Penicillin	10 units	≥ 29	-	≤ 28	<p>(a) Penicillin should be used to test the susceptibility of all staphylococci to all penicillinase-labile penicillins. Penicillin-resistant strains of staphylococci produce β-lactamase. Perform test(s) to detect β-lactamase production on staphylococci for which the penicillin MICs are ≤ 0.12 μg/mL or zone diameters ≥ 29 mm before reporting the isolate as penicillin susceptible. Rare isolates of staphylococci that contain genes for β-lactamase production may appear negative by β-lactamase tests. Consequently, for serious infections requiring penicillin therapy, laboratories should perform MIC tests and β-lactamase testing on all subsequent isolates from the same patient. PCR testing of the isolate for the <i>blaZ</i> β-lactamase gene may be considered. See Table 3F, Page 150-153.</p> <p>(b) For methicillin (oxacillin)-resistant staphylococci report penicillin as resistant or do not report.</p>



***Staphylococcus aureus* (continued)**

**PENICILLINASE-STABLE PENICILLINS**

<p>Oxacillin (Oxacillin disk testing is not reliable for <i>S. aureus</i> and <i>S. lugdunensis</i>.)</p>	<p>30 µg Cefoxitin (surrogate test for oxacillin)</p>	<p>≥ 22 (cefoxitin)</p>	<p>-</p>	<p>≤ 21 (cefoxitin)</p>	<p>(a) Cefoxitin is tested as a surrogate for oxacillin for some species of <i>Staphylococcus</i>. Isolates that test resistant by cefoxitin or oxacillin, when using the appropriate test method for the species, should be reported as methicillin (oxacillin) resistant. If testing only cefoxitin, report as methicillin (oxacillin) susceptible or resistant based on the cefoxitin result. Isolates that test either <i>mecA</i> negative or PBP2a negative or cefoxitin susceptible should be reported as methicillin (oxacillin) susceptible.</p> <p>(b) For isolates of <i>S.aureus</i> that do not grow well on CAMHB or unsupplemented MHA (eg, small-colony variants), testing on other media (eg, BMHA) does not reliably detect <i>mecA</i>-mediated resistance. Testing for PBP2a using induced growth (ie, growth taken from the zone margin surrounding a cefoxitin disk on either BMHA or a blood agar plate after 24 hours incubation in 5% CO<sub>2</sub>) or <i>mecA</i> should be done. *Cation Adjusted Mueller Hinton Agar</p>
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المنظمة الصحية  
المملكة العربية السعودية

<b><i>Staphylococcus aureus</i> (continued)</b>														
<b>GLYCOPEPTIDES</b>														
Vancomycin	-	-	-	-	<p>(a) For <i>S. aureus</i>, vancomycin-susceptible isolates may become vancomycin intermediate during the course of prolonged therapy.</p> <p>(b) MIC tests should be performed to determine the susceptibility of all isolates of staphylococci to vancomycin. The disk test does not differentiate vancomycin-susceptible isolates of <i>S. aureus</i> from vancomycin-intermediate isolates, nor does the test differentiate among vancomycin susceptible, -intermediate, and -resistant isolates of <i>Staphylococcus</i> spp. other than <i>S. aureus</i> all of which give similar size zones of inhibition.</p> <p>(c) Send any <i>S. aureus</i> for which the vancomycin is <math>\geq 8</math> <math>\mu\text{g/mL}</math> to a reference laboratory.</p> <table border="1"> <thead> <tr> <th colspan="3"><b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b></th> </tr> <tr> <th><b>S</b></th> <th><b>I</b></th> <th><b>R</b></th> </tr> </thead> <tbody> <tr> <td><math>\leq 2</math></td> <td>4-8</td> <td><math>\geq 16</math></td> </tr> </tbody> </table>	<b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b>			<b>S</b>	<b>I</b>	<b>R</b>	$\leq 2$	4-8	$\geq 16$
<b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b>														
<b>S</b>	<b>I</b>	<b>R</b>												
$\leq 2$	4-8	$\geq 16$												
Teicoplanin (Optional) (Investigation)	-	-	-	-	<table border="1"> <thead> <tr> <th colspan="3"><b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b></th> </tr> <tr> <th><b>S</b></th> <th><b>I</b></th> <th><b>R</b></th> </tr> </thead> <tbody> <tr> <td><math>\leq 8</math></td> <td>16</td> <td><math>\geq 32</math></td> </tr> </tbody> </table>	<b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b>			<b>S</b>	<b>I</b>	<b>R</b>	$\leq 8$	16	$\geq 32$
<b>Interpretive Categories and MIC Breakpoints, <math>\mu\text{g/mL}</math></b>														
<b>S</b>	<b>I</b>	<b>R</b>												
$\leq 8$	16	$\geq 32$												
<b>TETRACYCLINES</b>														
Doxycycline	30 $\mu\text{g}$	$\geq 16$	13-15	$\leq 12$										
<b>MACROLIDES</b>														
Erythromycin	15 $\mu\text{g}$	$\geq 23$	14-22	$\leq 13$	Not routinely reported on organisms isolated from the urinary tract.									
<b>FLUOROQUINOLONES</b>														
Ciprofloxacin	5 $\mu\text{g}$	$\geq 21$	16-20	$\leq 15$	<i>Staphylococcus</i> spp. may develop resistance during prolonged therapy with quinolones. Therefore, isolates that are initially susceptible may become resistant within three to four days after initiation of therapy. Testing of repeat isolates may be warranted.									





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<b><i>Staphylococcus aureus</i> (continued)</b>					
<b>NITROFURANTOINS</b>					
Nitrofurantoin	300 µg	≥ 17	15-16	≤ 14	For testing and reporting urinary tract isolates only
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 16	11-15	≤ 10	
<b>LINCOSAMIDES</b>					
Clindamycin	2 µg	≥ 21	15-20	≤ 14	<p><b>(a) Not routinely reported on organisms isolated from the urinary tract.</b></p> <p>(b) For isolates that test erythromycin resistant and clindamycin susceptible or intermediate, testing for ICR by disk diffusion using the D-zone test or by broth microdilution is required before reporting clindamycin (See Table 3I, Page 160-162).</p> <p>(c) D-zone test: 15-µg erythromycin and 2-µg clindamycin disks spaced 15–26 mm apart. Report isolates with ICR as "clindamycin resistant" (See Table 3I, Page 160-162).</p> <p>*ICR: Inducible clindamycin resistance</p>
<b>ANSAMYCINS</b>					
Rifampin	5 µg	≥ 20	17-19	≤ 16	<p>(a) Rifampin should be used but not reported.</p> <p>(b) Rx: should not be used alone for antimicrobial therapy.</p>



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<b><i>Enterococcus</i> spp.</b>					
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	I	R	
<b>PENICILLINS</b>					
Ampicillin	10 µg	≥ 17	-	≤ 16	The results of ampicillin susceptibility tests should be used to predict the activity of amoxicillin. Ampicillin results may be used to predict susceptibility to amoxicillin-clavulanate, ampicillin-sulbactam, and piperacillin-tazobactam among non-β-lactamase producing enterococci. Ampicillin susceptibility can be used to predict imipenem susceptibility, providing the species is confirmed to be <i>E. faecalis</i> .
<b>GLYCOPEPTIDES</b>					
Vancomycin	30 µg	≥ 17	15-16	≤ 14	When testing vancomycin against enterococci, plates should be held a full 24 hours for accurate detection of resistance. Zones should be examined using transmitted light; the presence of a haze or any growth within the zone of inhibition indicates resistance. Organisms with intermediate zones should be tested by an MIC method as described in M07. For isolates for which the vancomycin MICs are 8 to 16 µg/mL, perform biochemical tests for identification as listed under the “Vancomycin MIC ≥ 8 µg/mL” test found in Table 3H, Page 158-159.
<b>FLUOROQUINOLONES</b>					
Ciprofloxacin	5 µg	≥ 21	16-20 <sup>^</sup>	≤ 15	<b>For testing and reporting urinary tract isolates only.</b>
<b>NITROFURANTOINS</b>					
Nitrofurantoin	300 µg	≥ 17	15-16	≤ 14	For testing and reporting urinary tract isolates only.
<b>OXAZOLIDINONES</b>					
Linezolid	30 µg	≥ 23	21-22	≤ 20	

**Test for Gentamicin High-Level Aminoglycoside Resistance in *Enterococcus* spp.**

Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments
		S	Inconclusive	R	
Gentamicin	120 µg	≥ 10	7-9	= 6	If disk diffusion result is inconclusive: perform an agar dilution or broth dilution MIC test to confirm (See Table 3K, Page 166-168).



\* For disk diffusion, test a maximum of 9 disks on a 150-mm plate and 4 disks on a 100-mm plate.

<b><i>Streptococcus pneumoniae</i></b>							
Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Comments		
		S	I	R			
<b>PENICILLINS</b>							
Penicillin (nonmeningitis)	1 µg Oxacillin	≥ 20	-	-	Isolates of pneumococci with oxacillin zone sizes of ≥20 mm are susceptible (MIC ≤ 0.06 µg/mL) to penicillin. Penicillin and cefotaxime, ceftriaxone, or meropenem MICs should be determined for those isolates with oxacillin zone diameters of ≤ 19 mm, because zones of ≤ 19 mm occur with penicillin-resistant, -intermediate, or certain -susceptible strains. For isolates with oxacillin zones ≤ 19 mm, do not report penicillin as resistant without performing a penicillin MIC test.		
Penicillin parenteral (nonmeningitis) (optional)	-	-	-	-	<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>		
					<b>S</b>	<b>I</b>	<b>R</b>
					≤ 2	4	≥ 8
Rx: Doses of intravenous penicillin of at least 2 million units every 4 hours in adults with normal renal function (12 million units per day) can be used to treat nonmeningeal pneumococcal infections due to strains with penicillin MICs ≤ 2 µg/mL. Strains with an intermediate MIC of 4 µg/mL may require penicillin doses of 18 to 24 million units per day.							
<b>CEPHEMS</b>							
Ceftriaxone (nonmeningitis)	-	-	-	-	<b>Interpretive Categories and MIC Breakpoints, µg/mL</b>		
					<b>S</b>	<b>I</b>	<b>R</b>
					≤ 1	2	≥ 4
<b>TETRACYCLINES</b>							
Doxycycline	30 µg	≥ 28	25-27	≤ 24	Organisms that are susceptible to tetracycline are also considered susceptible to doxycycline. However, resistance to doxycycline cannot be inferred from tetracycline resistance.		



<b><i>Streptococcus pneumoniae</i> (continued)</b>					
<b>MACROLIDES</b>					
Erythromycin	15 µg	≥ 21	16-20	≤ 15	(a) Susceptibility and resistance to azithromycin, clarithromycin, and dirithromycin can be predicted by testing erythromycin.  (b) Not routinely reported on organisms isolated from the urinary tract.
<b>FLUOROQUINOLONES</b>					
Levofloxacin	5 µg	≥ 17	14-16	≤ 13	
<b>FOLATE PATHWAY INHIBITORS</b>					
Trimethoprim-sulfamethoxazole	1.25/ 23.75 µg	≥ 19	16-18	≤ 15	
<b>LINCOSAMIDES</b>					
Clindamycin	2 µg	≥ 19	16-18	≤ 15	<b>(a) Not routinely reported on organisms isolated from the urinary tract.</b> (b) For isolates that test erythromycin resistant and clindamycin susceptible or intermediate, testing for ICR by disk diffusion using the D-zone test or by broth microdilution is required before reporting clindamycin (See Table 3I, Page 160-162). (c) D-zone test: 15-µg erythromycin and 2-µg clindamycin disks spaced 15–26 mm apart. Report isolates with ICR as "clindamycin resistant" (See Table 3I, Page 160-162). *ICR: Inducible clindamycin resistance

Note: Information in boldface type is new or modified since the previous edition.

\*Intermediate ranges denoted with a "^" for the applicable antimicrobial agents in the drug groups are based on the known ability of these agents to concentrate in the urine.